

# Determining Food Safety Best Practices for Pecan Float Systems

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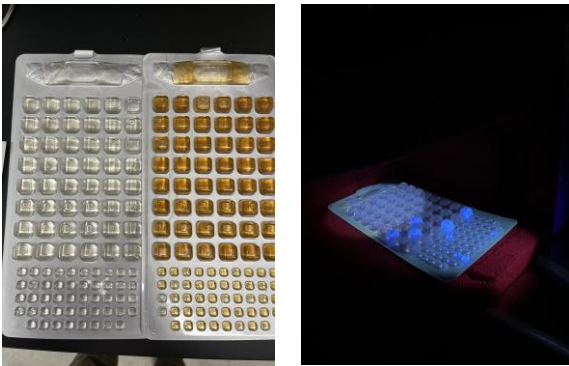
# Pecan Float Systems Overview

- In order to optimize kernel yield, pecans will go through a float system to separate shells from the kernels.
- Generally chemical sanitizers are used to maintain water quality
- Sanitizer levels may be difficult to maintain as the organic load through recirculated water systems increase.
- If not properly handled, water can be a potential source of cross-contamination of foodborne pathogens in a recirculated system.

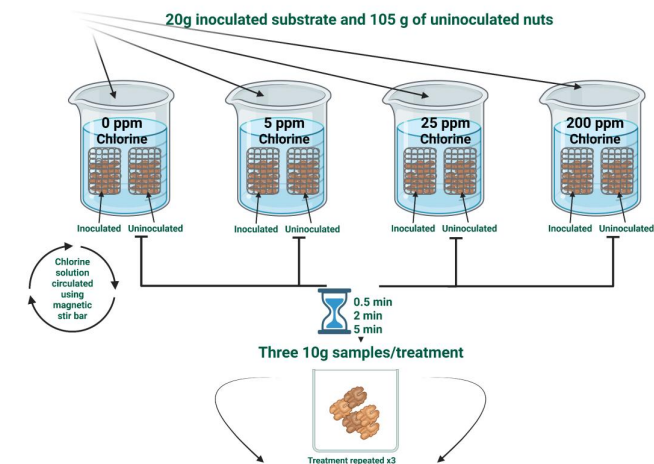
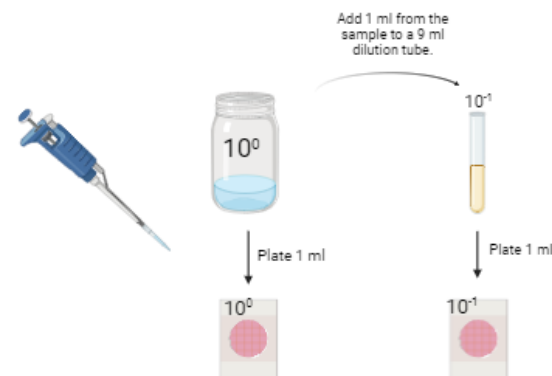


# Objectives

- Evaluate the physiochemical and microbial quality of pecan float systems.
- Evaluate how interventions may impact and improve the microbial quality of the water and pecan kernels in a float system.
- Determine the effective minimum free chlorine levels that prevent the cross-contamination of *Salmonella* in a simulated pecan float system.



Water *E. coli*/Coliform Petrifilm Plating



# Float Tank Microbial and Physiochemical Quality (Methods)

## Interventions

**Baseline:** Samples taken pre-intervention.

**Intervention 1:** Float tank water spiked with chlorine every 60 minutes.

**Intervention 2:** Peracetic acid spray bar at the end of the process.

## Physiochemical Analysis



### **Colorimeter**

- Total Chlorine (ppm)
- Free Chlorine (ppm)
- Chemical Oxygen Demand (COD, ppm)

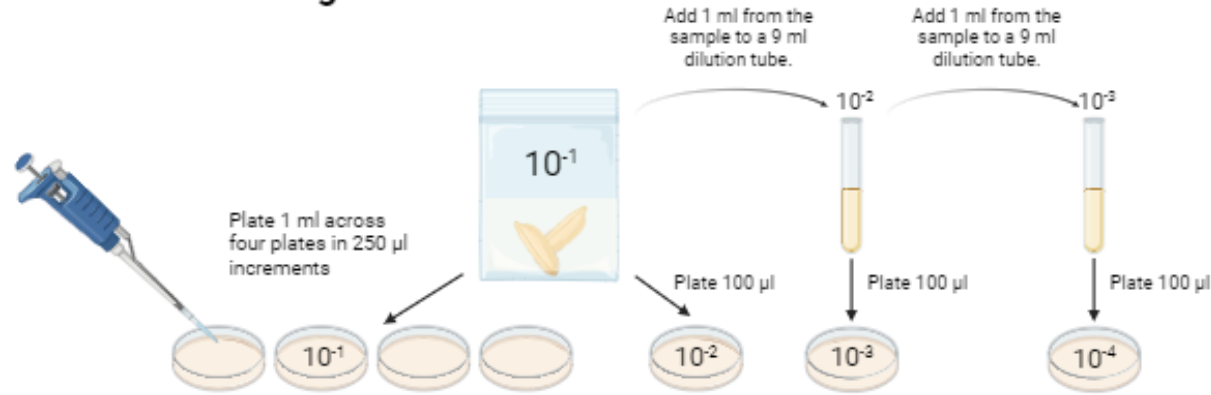


### **pH Meter**

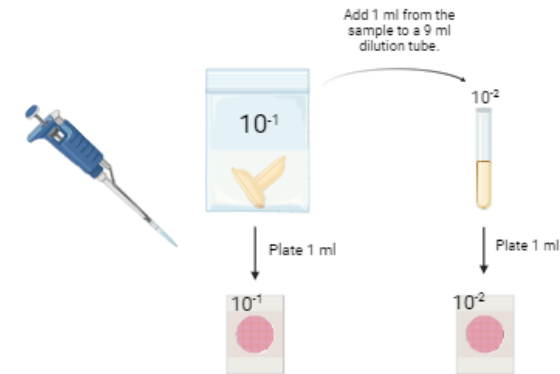
# Float Tank Microbial and Physiochemical Quality (Methods)

## Microbial Analysis of Water and Pecan Kernels

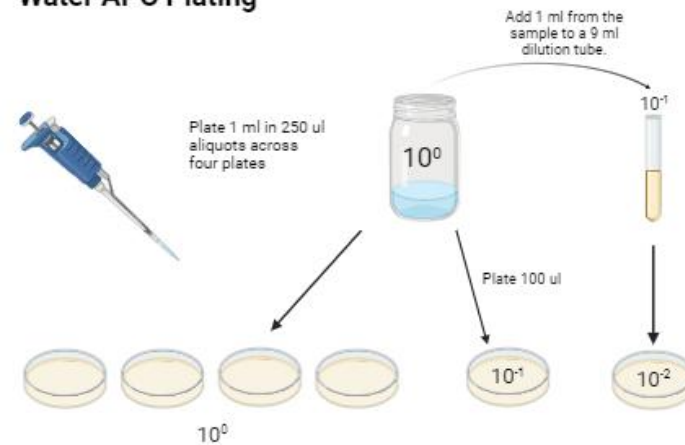
### Pecan APC Plating



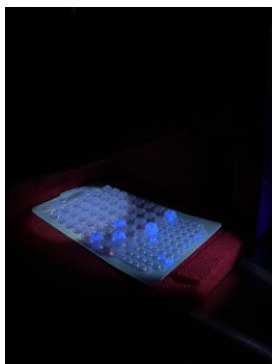
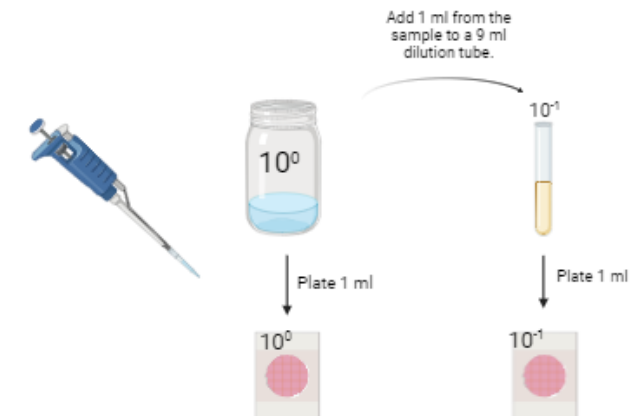
### Pecan *E. coli*/Coliform Petrifilm Plating



### Water APC Plating

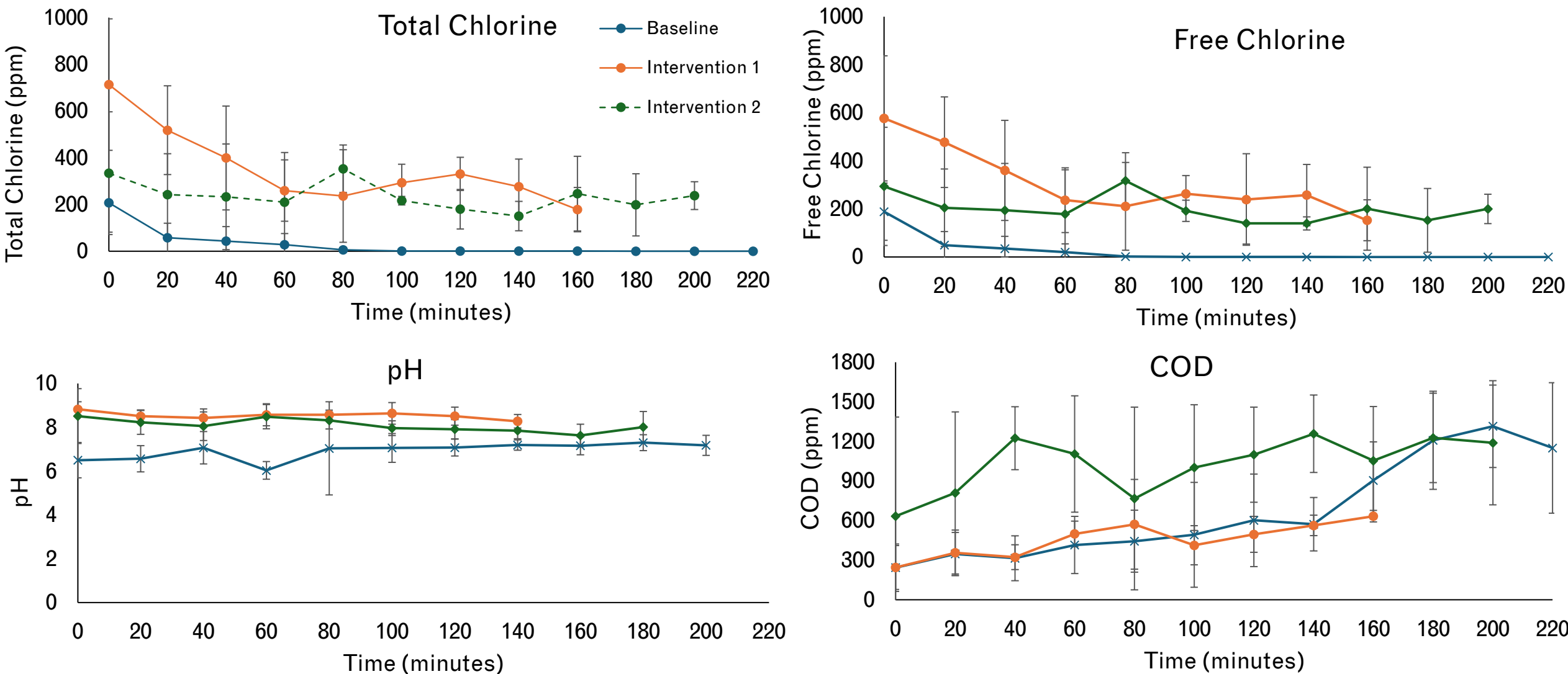


### Water *E. coli*/Coliform Petrifilm Plating



# Float Tank Microbial and Physiochemical Quality (Results)

**Figure 3.** Average Total chlorine, free chlorine, pH, and COD measurements in the float system water for each intervention over the processing period (n=3)



# Float Tank Microbial and Physiochemical Quality (Results)

**Table 3.** Average  $\pm$  standard deviation APC and coliform counts of water samples taken from the float system for each intervention over the course of the processing period (n=3).

Timepoint (Minutes)	APC (log CFU or MPN/100 ml)			Coliform (log CFU or MPN/100 ml)		
	Baseline	Inter. 1	Inter. 2	Baseline	Inter. 1	Inter. 2
0	0.87 $\pm$ 1.25	0.16 $\pm$ 0.28	1.13 $\pm$ 1.95	0.00 $\pm$ 0.00	0.00 $\pm$ 0.00	0.00 $\pm$ 0.00
20	2.31 $\pm$ 0.73	0.00 $\pm$ 0.00	1.80 $\pm$ 1.33	1.69 $\pm$ 1.47	0.00 $\pm$ 0.00	0.00 $\pm$ 0.00
40	2.74 $\pm$ 0.92	0.80 $\pm$ 0.73	1.98 $\pm$ 1.53	1.56 $\pm$ 0.77	0.00 $\pm$ 0.00	0.00 $\pm$ 0.00
60	2.97 $\pm$ 0.88	0.98 $\pm$ 0.84	2.20 $\pm$ 1.73	1.88 $\pm$ 1.45	0.00 $\pm$ 0.00	0.00 $\pm$ 0.00
80	3.32 $\pm$ 1.39	1.52 $\pm$ 2.38	0.72 $\pm$ 0.66	2.10 $\pm$ 1.88	0.00 $\pm$ 0.00	0.00 $\pm$ 0.00
100	3.43 $\pm$ 1.49	0.36 $\pm$ 0.32	1.41 $\pm$ 0.21	2.74 $\pm$ 1.93	0.00 $\pm$ 0.00	0.00 $\pm$ 0.00
120	4.24 $\pm$ 0.78	0.00 $\pm$ 0.00	1.59 $\pm$ 0.37	3.75 $\pm$ 0.93	0.00 $\pm$ 0.00	0.00 $\pm$ 0.00
140	3.41 $\pm$ 2.09	0.53 $\pm$ 0.46	1.57 $\pm$ 0.75	3.68 $\pm$ 0.99	0.00 $\pm$ 0.00	0.00 $\pm$ 0.00
160	4.12 $\pm$ 0.89	1.55 $\pm$ 1.39	1.44 $\pm$ 1.30	3.40 $\pm$ 0.43	0.00 $\pm$ 0.00	0.00 $\pm$ 0.00
180	4.52 $\pm$ 1.19	NA	1.24 $\pm$ 1.13	3.83 $\pm$ 1.01	NA	0.00 $\pm$ 0.00
200	4.53 $\pm$ 1.32	NA	1.33 $\pm$ 0.42	3.68 $\pm$ 1.19	NA	0.00 $\pm$ 0.00
220	4.51 $\pm$ 0.42	NA	NA	3.23 $\pm$ 0.64	NA	NA

# Float Tank Microbial and Physiochemical Quality (Results)

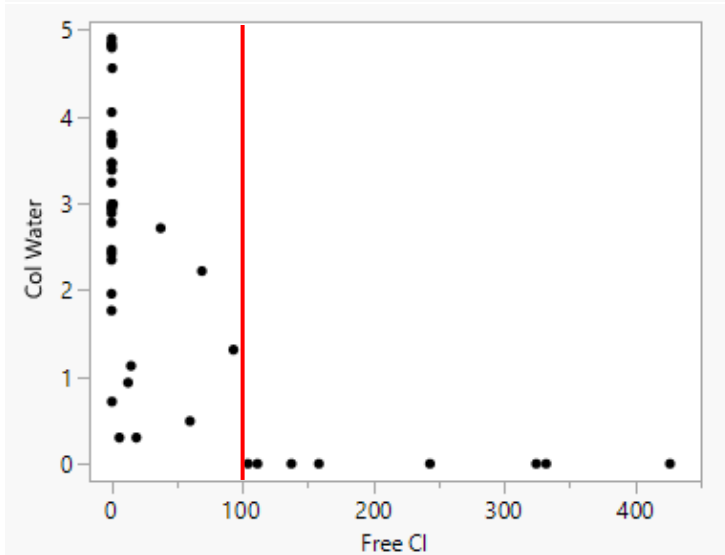
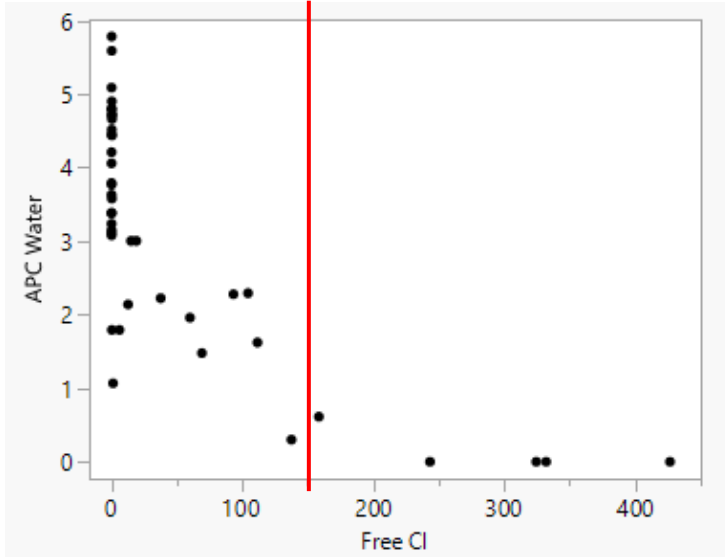
**Table 4.** Pearson correlation table of the physiochemical and microbial water quality parameters of the pecan float tank water ( $p < 0.05$ )

	APC	Col	Tot Cl 2	Free Cl 2	pH	COD
APC	1.000	0.808	-0.783	-0.767	-0.611	0.307
Col	0.808	1.000	-0.596	-0.573	-0.509	0.001
Tot Cl 2	-0.783	-0.596	1.000	0.984	0.752	-0.312
Free Cl 2	-0.767	-0.573	0.984	1.000	0.744	-0.330
pH	-0.611	-0.509	0.752	0.744	1.000	-0.181
COD	0.307	0.001	-0.312	-0.330	-0.181	1.000

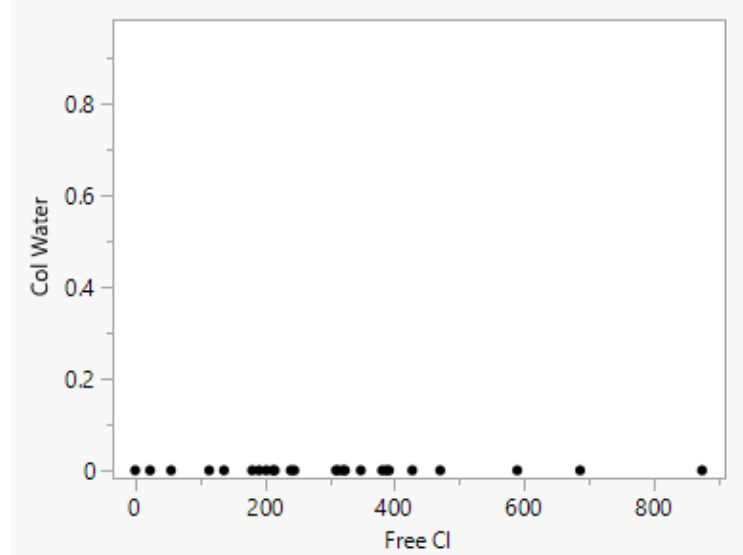
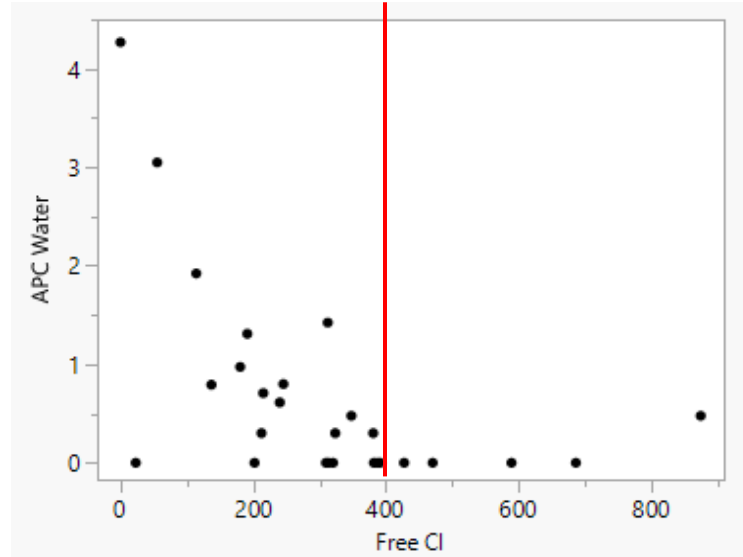


# Float Tank Microbial and Physiochemical Quality (Results)

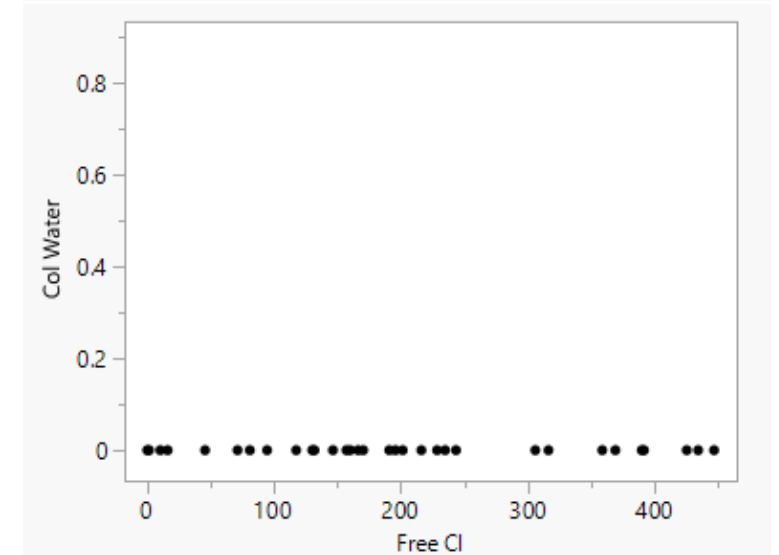
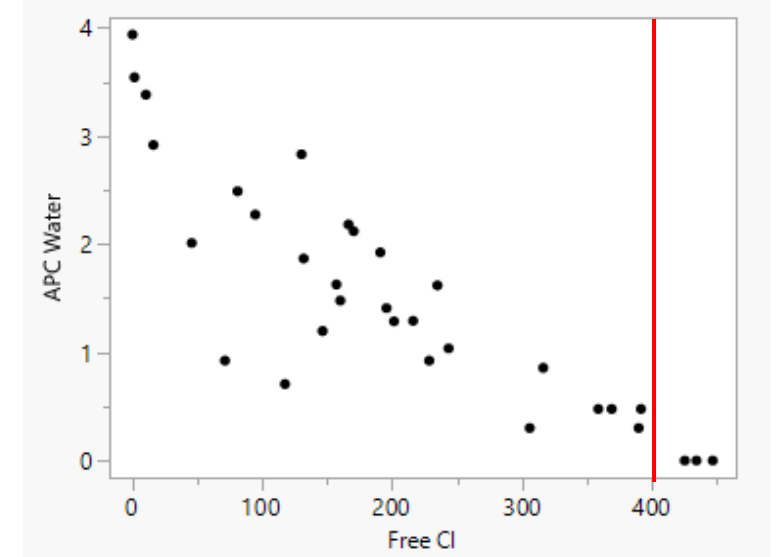
Baseline



Intervention 1

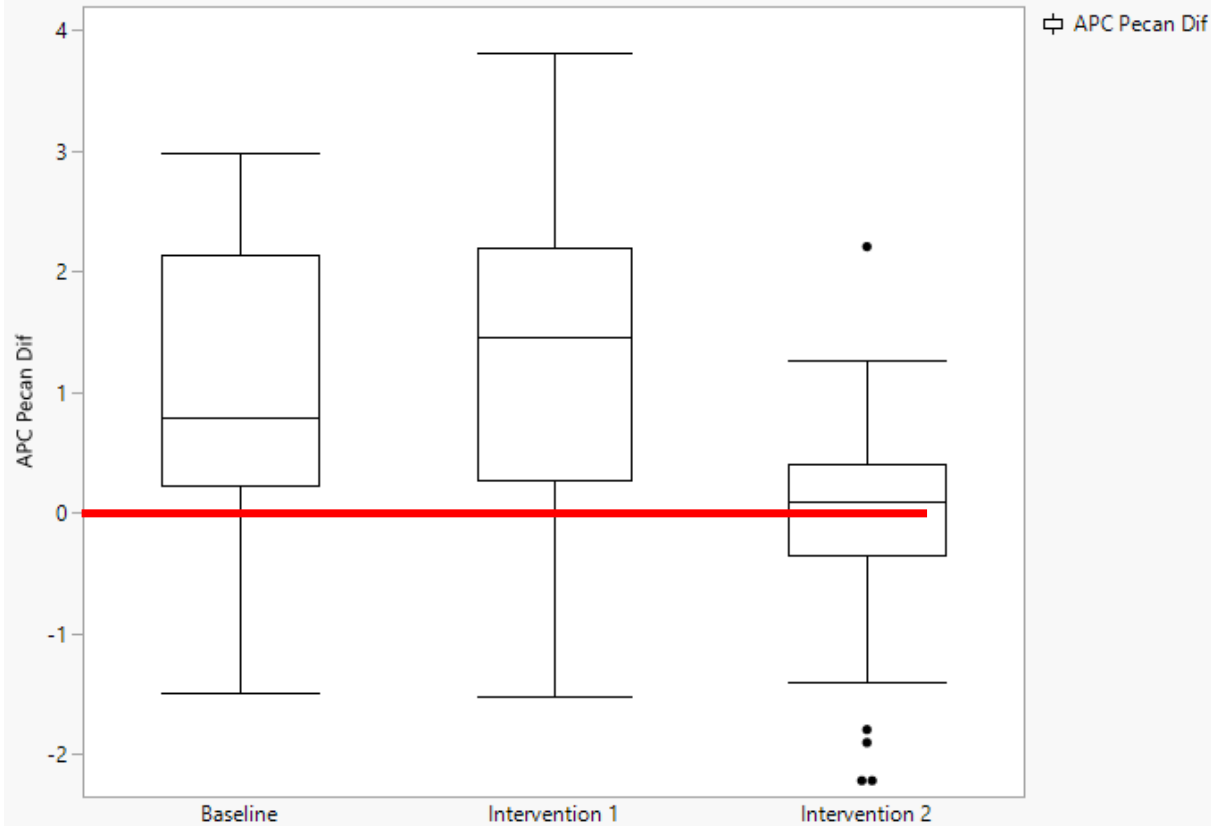


Intervention 2

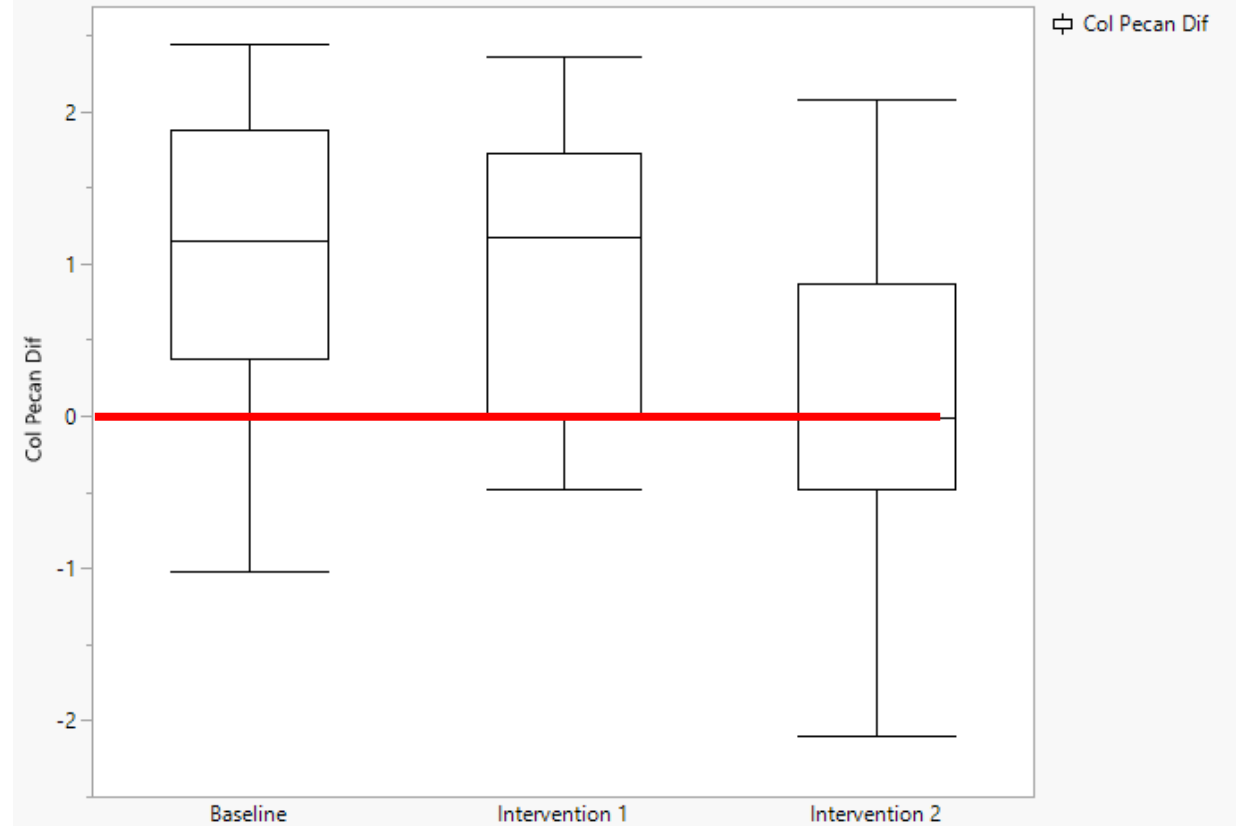


# Float Tank Microbial and Physiochemical Quality (Results)

APC Pecan Dif vs. Intervention



Col Pecan Dif vs. Intervention



# Minimum Chlorine Level to Prevent Cross-Contamination (Methods)

Overnight Bacterial lawn from TSA +R plate flooded with 250ul rifampicin resistant S. Typhimurium, Enteritidis PT 9c, Montevideo, Saintpaul, or Newport

Lawn plates flooded with 5mL 0.1% peptone water, scraped, and collected into single falcon tube

Inoculated substrate placed under biocontainment hood until visibly dry

100/0, 50/50, or 25/75% shell/halves

10mL inoculum/100g substrate

Alternately massaged and shaken

01:30:00

10mL 0.1% peptone water

1:2 Dilution

Spread plated on to Tryptic Soy Agar supplemented with rifampicin

Incubated @ 37°C for 24h and enumerated

Serial dilution in 0.1% peptone water

Homogenized for 30 seconds

20g inoculated substrate and 105 g of uninoculated nuts

0 ppm Chlorine

Inoculated Uninoculated

5 ppm Chlorine

Inoculated Uninoculated

25 ppm Chlorine

Inoculated Uninoculated

200 ppm Chlorine

Inoculated Uninoculated

Chlorine solution circulated using magnetic stir bar

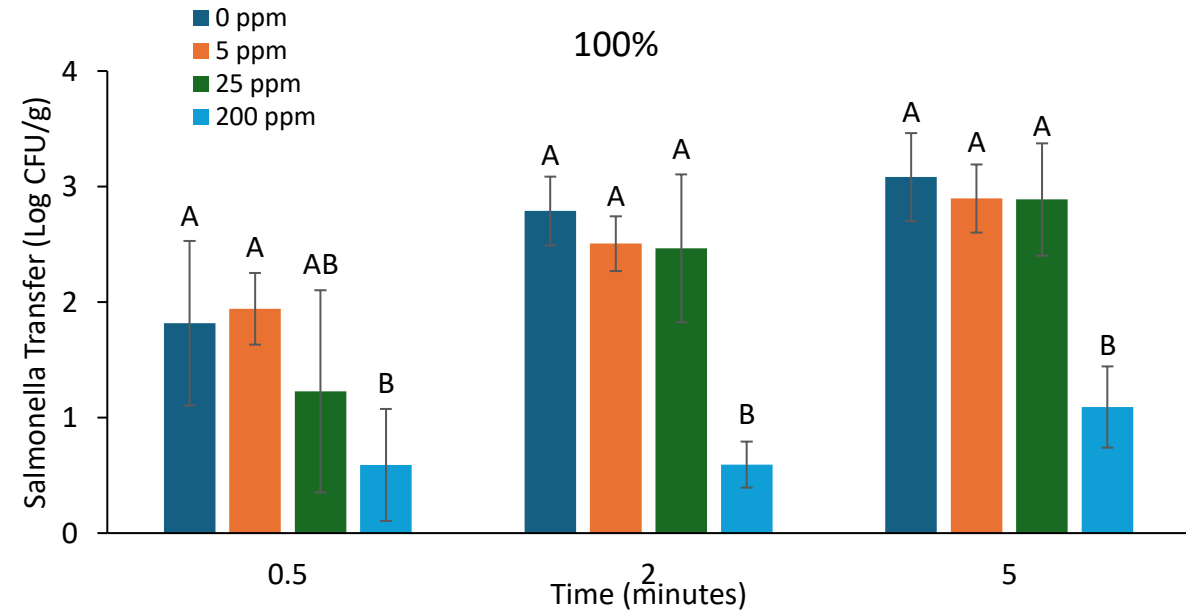
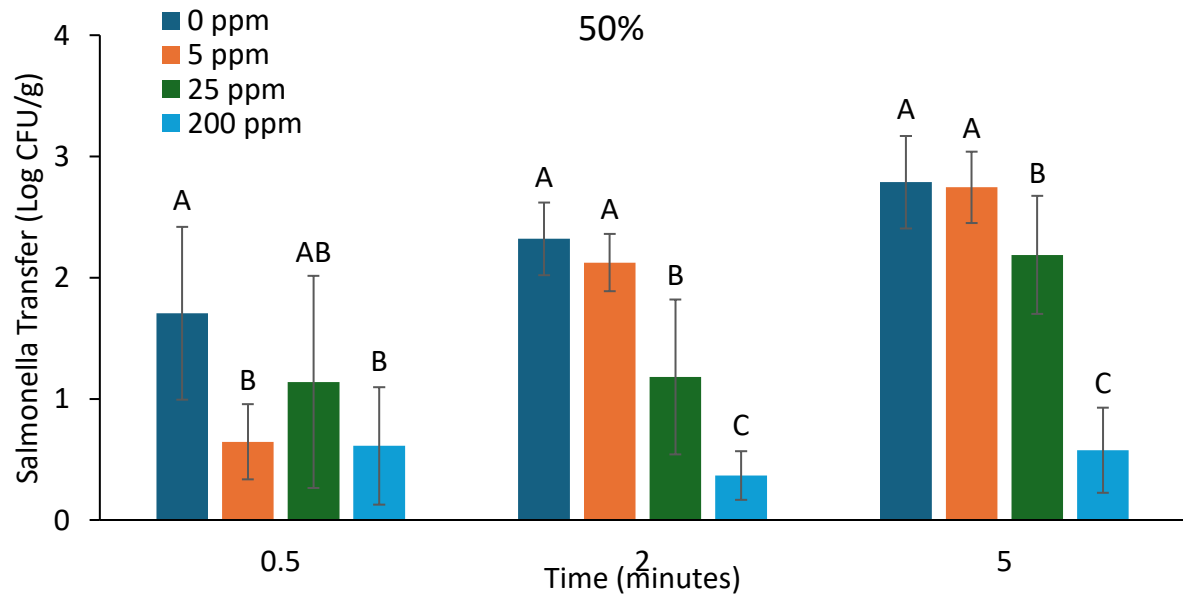
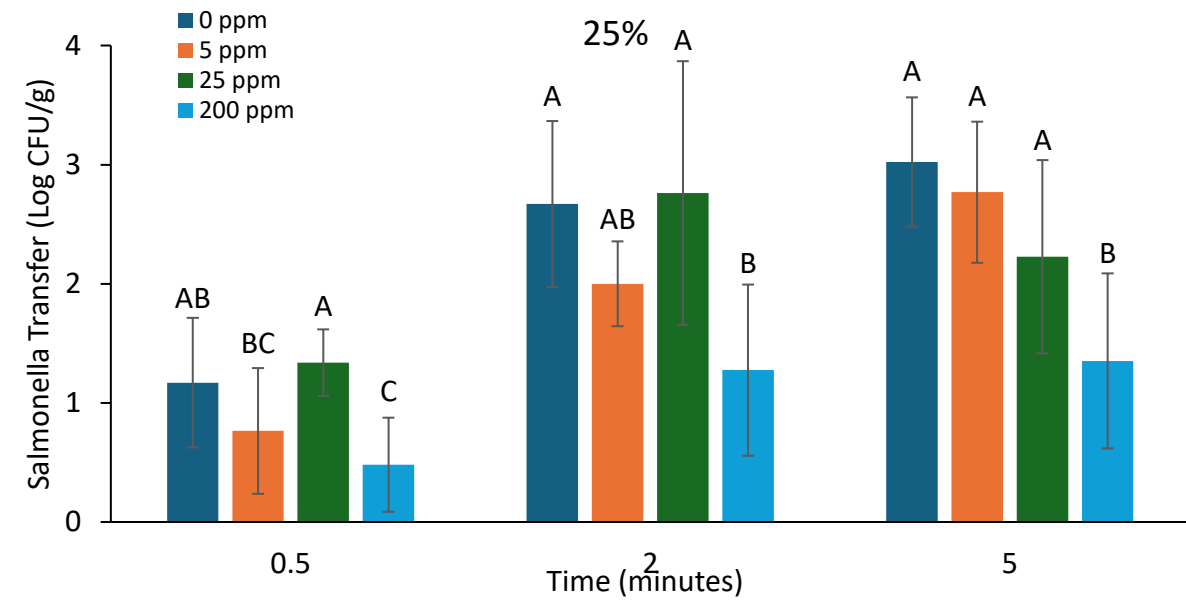
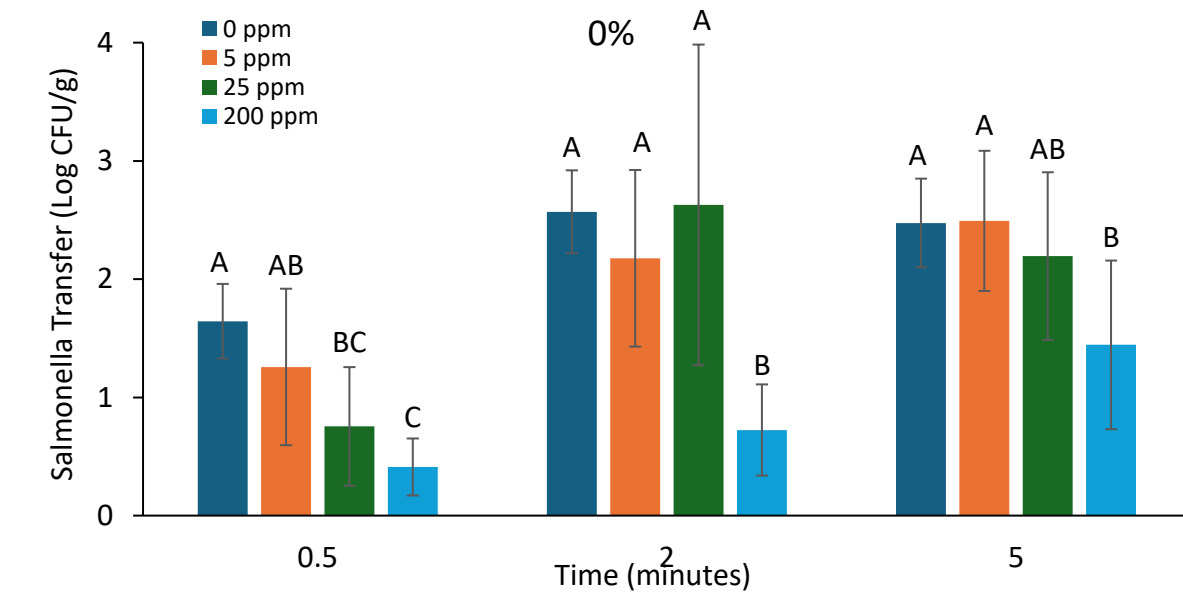
0.5 min  
2 min  
5 min

Three 10g samples/treatment

Treatment repeated x3

Data was analyzed by General Linear Model and Tukey's HSD ( $P \leq 0.05$ ).

# Minimum Chlorine Level to Prevent Cross-Contamination (Results)



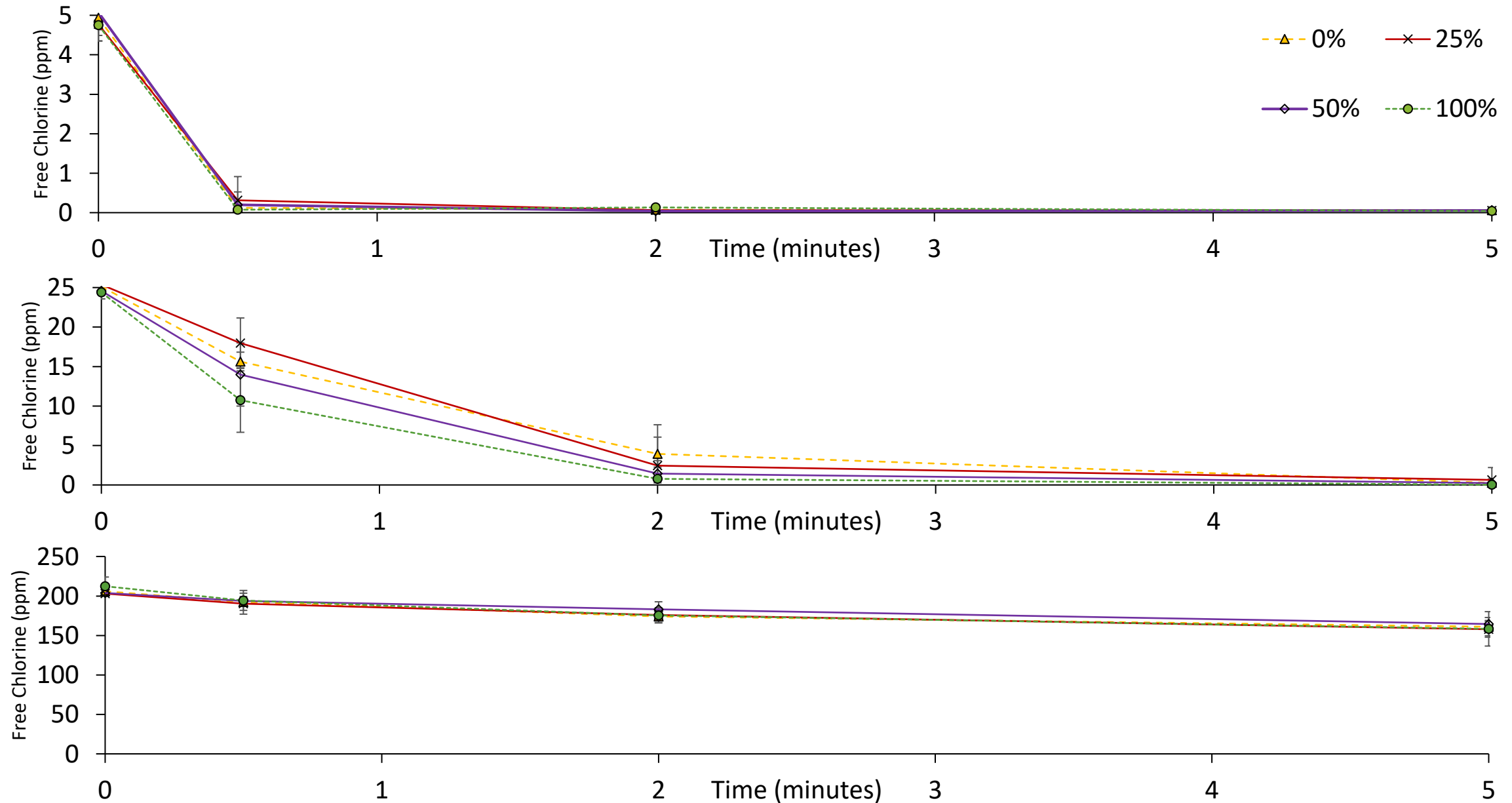


# Minimum Chlorine Level to Prevent Cross-Contamination (Results)

Table 1. *Salmonella* transfer to uninoculated pecan samples in a simulated float tank at 0, 5, 25, and 200 ppm free chlorine levels at 0.5, 2, and 5 minutes with 0, 25, 50, and 100% inoculated kernel-shell ratio.

Time (minutes)	0 ppm A				5 ppm A				25 ppm B				200 ppm C			
	0%	25%	50%	100%	0%	25%	50%	100%	0%	25%	50%	100%	0%	25%	50%	100%
0.5 A	(4/9)	(4/9)	(7/9)	(6/9)	(5/9)	(3/9)	(4/9)	(7/9)	(0/9)	(4/9)	(4/9)	(4/9)	(0/9)	(3/9)	(4/9)	(2/9)
2 B	(9/9)	(9/9)	(9/9)	(9/9)	(9/9)	(8/9)	(8/9)	(9/9)	(7/9)	(8/9)	(5/9)	(8/9)	(8/9)	(9/9)	(4/9)	(0/9)
5 B	(9/9)	(9/9)	(9/9)	(9/9)	(8/9)	(8/9)	(9/9)	(9/9)	(4/9)	(9/9)	(9/9)	(8/9)	(4/9)	(1/9)	(4/9)	(2/9)

# Minimum Chlorine Level to Prevent Cross-Contamination (Results)



# Key Takeaways

- Once free chlorine levels were below 200 ppm aerobic plate counts (microbial load) increased and once below 100 ppm coliform counts (indicator organisms) increased in the float water.
- Intervention 1 (chlorine spike every 60 minutes) improved the microbial quality of the water and eliminated the coliforms in the float water.
- Intervention 2 (peracetic spray bar at the end of the system) significantly improved the microbial quality of the pecans after going through the float system.
- Increased time in the float system significantly impacted the transfer of *Salmonella* to the kernels, except for the 200 ppm free chlorine treatment
- Free chlorine levels around 200 ppm free chlorine were most effective at preventing cross-contamination of *Salmonella*.



# Future Research

- Optimize in-line pulsed light treatments to maximize reductions.
- Determine optimal hot water sanitation parameters to achieve a 5-log reduction while maintaining quality and improve shelling.
- Evaluate the antimicrobial properties of coatings and modified packaging.
- Investigate the effectiveness of gas treatments on foodborne pathogens while maintaining quality.
  - Ozone
  - Chlorine Dioxide
- Evaluate promising end of process pathogen treatments.
  - E-Beam/Irradiation
  - Pulsed Electric Field
- Determine controls and best practices to reduce contamination along the supply chain.



# Questions



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